

# Antimicrobial effects of tea-polyphenols on pathogenic *Vibrio parahaemolyticus* in shucked oysters (*Crassostrea plicatula*)

X. Shen<sup>1,2</sup>, C. Liu<sup>2\*</sup>, and Y. Su<sup>3\*</sup>

<sup>1</sup> East China Sea Fisheries Research Institute, Chinese Fisheries Academy of Fishery Science, 300 Jungong Road, Shanghai 200090, China (foodsmc98@126.com)

<sup>2</sup> College of Food Science and Technology, Shanghai Ocean University, 999 Huchenghuan Road, Shanghai 201306, China ([chengchuliu@yahoo.com](mailto:chengchuliu@yahoo.com))

<sup>3</sup> Seafood Research and Education Center, Oregon State University, Astoria, Oregon 97103, USA ([yi-cheng.su@oregonstate.edu](mailto:yi-cheng.su@oregonstate.edu))

\*corresponding authors

## Introduction

*Vibrio parahaemolyticus* is a Gram-negative human pathogen that is naturally present in inshore marine waters (Cook *et al.*, 2002) and recognized as the leading causes of foodborne gastroenteritis associated with seafood consumption throughout the world (Joseph *et al.*, 1982; Su and Liu, 2007). Tea-polyphenols are natural edible compounds extracted from tea leaves with excellent antimicrobial activity against food spoilage and foodborne pathogens including *Staphylococcus aureus*, *Escherichia coli*, *Streptococcus mutans*, *Mycobacterium tuberculosis*, *Bacillus cereus* and *Listeria monocytogenes* (An *et al.*, 2004; Paras *et al.*, 2006; Si *et al.*, 2006; Pilar *et al.*, 2008). However, there is no information about tea polyphenols against growth of pathogenic *V. parahaemolyticus*. This study investigated the inhibitory effects of tea-polyphenols against *V. parahaemolyticus* in both culture suspension and contaminated shucked oyster and provided useful information for improving safety of oysters for consumption.

## Materials and Methods

### ***Bacteria strains and preparation of multi-strain cocktail suspension***

Two virulent strains *V. parahaemolyticus* possessing *tdh* gene (ATCC 33846 and ATCC 33847), and one non-virulent strain (VP 1602 isolated from oysters) were used and a multi-strain cocktail culture suspension was prepared according to Shen *et al.* (2009).

### ***Growth inhibitory test of tea-polyphenols against V. parahaemolyticus***

The minimal inhibitory concentration (MIC) of tea-polyphenols against *V. parahaemolyticus* was determined according to An *et al.* (2004).

### ***Inoculation of V. parahaemolyticus in oysters***

Live *C. plicatula* oysters were collected and inoculated with *V. parahaemolyticus* by holding oysters in artificial seawater containing *V. parahaemolyticus* ( $10^4$ - $10^5$ CFU/mL) with circulating air at 10-12 °C for 4 h (Shen *et al.*, 2009)

### ***Effects of tea-polyphenols on V. parahaemolyticus in shucked oysters***

The inoculated oysters were shucked in a disinfected circumstance and then dipped in various tea-polyphenols solutions for 2.5 h. The populations of *V. parahaemolyticus* in shucked oysters were detected every 30 min. For storage experiment, the shucked oysters were treated with and without tea-polyphenols (0.5 mg/mL) and stored at 0°C, 5°C, and 10°C for 60 h respectively. The populations of *V. parahaemolyticus* in shucked oysters were determined every 12 h using the FDA's Bacteriological Analytic Manual.

### ***Sensory test of shucked oysters with and without the treatment of tea-polyphenols***

The taste and flavor of raw and cooked oysters treated with and without tea-polyphenols were evaluated and scores were given to each sample (1: like; -1: unlike; 0: acceptable) by a test panel with 12 students from the lab.

## **Results**

### ***Growth inhibitory test of tea polyphenols against V. parahaemolyticus in culture suspension and shucked oysters***

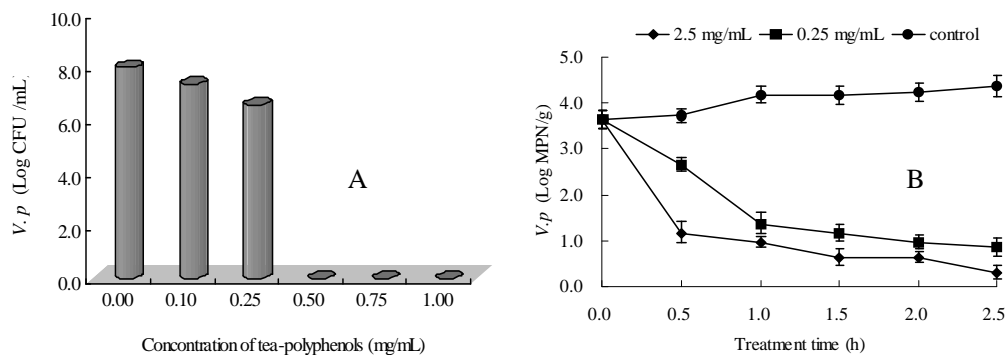


Figure 1 Inhibitory effect of tea-polyphenols against *V. parahaemolyticus* in culture broth (A) and shucked oysters (B)

Figure 1 (A) showed that the growth of *V. parahaemolyticus* in alkaline peptone water supplemented with 1.5% NaCl (APW-salt) at 37°C for 48 h was effectively inhibited by tea polyphenols. The MIC of tea-polyphenols against *V. parahaemolyticus* was 0.5 mg/mL. Figure 1 (B) demonstrated that the populations of *V. parahaemolyticus* in shucked oysters decreased greatly after the samples were treated with 0.25 and 2.5 µg/mL of tea polyphenols solutions. However, the population of *V. parahaemolyticus* in shucked oysters without treatment of tea-polyphenols was steadily increased. These results indicated the efficacy of tea-polyphenols to decontaminate *V. parahaemolyticus* in shucked oysters was correlated to both the treatment time and concentration of tea-polyphenols.

### ***Efficacy of tea polyphenols in decontamination of V. parahaemolyticus in shucked oysters during cold storage at different temperatures***

Figure 2 cold storage, no matter the storage temperatures were at 0, 5 or 10°C could effectively reduce the *V. parahaemolyticus* count in shucked oysters (Figure 2). However, *V. parahaemolyticus* reduction was much greater when the oysters were treated with tea-polyphenols before storage, indicating that tea-polyphenols could enhance the inhibitory effects of cold storage against *V. parahaemolyticus* in shucked oysters.

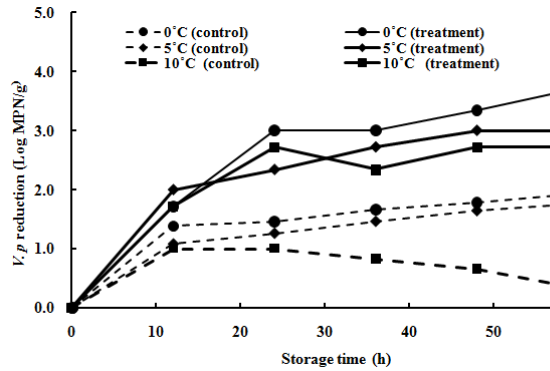


Figure 2 *V. parahaemolyticus* reduction in shucked oysters during storage with and without the treatment of tea-polyphenols

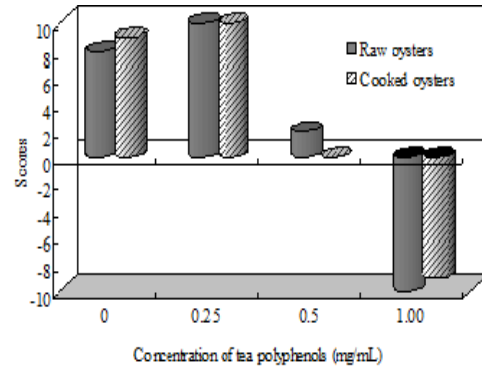


Figure 3 Effects of tea-polyphenols on the taste of raw and cooked oysters

### *Sensory acceptance of shucked oysters after the treatment of tea-polyphenols*

Figure 3 shows that the acceptance of oysters was dependent on the concentration of tea-polyphenols. The panel like the oysters treated by 0.25 mg/mL of tea-polyphenols, but did not like the oysters if the concentration of tea-polyphenols reached 1.00 mg/mL.

## Conclusions

Tea polyphenols had bactericidal effects against *V. parahaemolyticus* in both culture suspension and shucked oysters. Its minimal inhibition concentration was 0.50 mg/mL. The optimal concentration of tea polyphenols to treat shucked oysters was 0.25-0.5 mg/mL based on both antimicrobial and sensory tests. Those results indicated that tea polyphenols might be applied to shucked oysters to reduce the contamination of *V. parahaemolyticus*.

## Acknowledgements

This study was supported by Innovative Research Team Project of Universities and Leading Academic Discipline Project of Shanghai Municipal Education Commission (J50704).

## References

- An, B., Kwak, J., Son, J., Park, J., Lee, J., Jo, C., Byun, M. (2004) Biological and anti-microbial activity of irradiated green tea polyphenols. *Food Chemistry*. 88, 549-555.
- Cook, D.W., Leary, P.O., Hunsucker, J.C., Sloan, E.M., Bowers, J.C., Blodgett, R.J., DePaola, A. (2002) *Vibrio vulnificus* and *Vibrio parahaemolyticus* in U.S. retail shell oysters. *J. Food. Prot.* 65, 79-87.
- Paras, K., Deepak, K., Meera, S. (2006) Green tea polyphenol inhibits *Mycobacterium tuberculosis* survival within human macrophages. *INT J BIOCHEM CELL B I* 38, 600-609.
- Pilar, M., Rosa, C., Angel, J., Michael, H. (2008) Antioxidant and antimicrobial activities of tea infusions. *Food Chemistry* 108, 55-63.
- Shen, X., Cai, Y., Liu, C., Liu, W., Hui, Y., Su, Y. C. (2009) Effect of temperature on uptake and survival of *Vibrio parahaemolyticus* in oysters (*Crassostrea plicatula*). *Int J Food Microbiol* 136, 129-132.
- Si, W., Gong, J., Rong, T., Molosh, K., Raymond, Y., Yin, Y., 2006. Bioassay-guided purification and identification of antimicrobial components in Chinese tea extract. *J Chrom* 1125, 204-210.
- Su, Y.C. and Liu, C., 2007. *Vibrio parahaemolyticus*: A concern of seafood safety. *Food Microbiol.* 24, 549-558.